

1 **Endangered Australian top predator is frequently exposed to anticoagulant**
2 **rodenticides**

3 **ABSTRACT**

4 Anticoagulant rodenticides (ARs) used to control mammalian pest populations cause secondary
5 exposure of predatory species throughout much of the world. It is important to understand the
6 drivers of non-target AR exposure patterns as context for assessing long-term effects and
7 developing effective mitigation for these toxicants. In Australia, however, little is known about
8 exposure and effects of ARs on predators. We detected AR residues in 74% of 50
9 opportunistically collected carcasses of the Tasmanian wedge-tailed eagle (*Aquila audax fleayi*),
10 an endangered apex predator. In 22% of birds tested, or 31% of those exposed, liver
11 concentrations of second generation ARs (SGARs) were > 0.1 mg/kg ww. Eagles were exposed
12 to flocoumafen, a toxicant only available from agricultural suppliers, at an exceptionally high
13 rate (40% of birds tested). Liver SGAR concentrations were positively associated with the
14 proportion of agricultural habitat and human population density in the area around where each
15 eagle died. The high exposure rate, in a species not known to regularly prey upon synanthropic
16 rodents, supports the hypothesis that apex predators are vulnerable to SGARs. Our results
17 indicate that AR exposure constitutes a previously unrecognized threat to Tasmanian wedge-
18 tailed eagles and highlight the importance of efforts to address non-target AR exposure in
19 Australia.

20 **Key words:** Rodenticide, Environmental contamination, SGARs, Secondary poisoning, Predator

21 **1. Introduction**

22 Anticoagulant rodenticides (ARs) are used worldwide to control mammalian pest
23 populations. These compounds function by inhibiting blood clotting mechanisms in vertebrates,
24 resulting in internal hemorrhaging (Rattner et al., 2014). The discovery of resistance to the first-
25 generation of ARs (FGARs) led to the development of second-generation ARs (SGARs) in the
26 1970s (van den Brink et al., 2018). To be lethal, FGARs generally require consecutive intake
27 over several days to accumulate sufficiently high concentrations (Erickson and Urban, 2004).
28 Conversely, SGARs are usually lethal from a single exposure and persist longer in the
29 environment (Erickson and Urban, 2004; van den Brink et al., 2018). The persistence of AR
30 compounds (Horak et al., 2018), the delay in mortality after bait consumption (Lee et al., 2006)
31 and the behavioral changes that occur as a symptom of poisoning (Brakes and Smith, 2005;
32 Mooney, 2017) can make poisoned rodents AR vectors to non-target predatory species.

33 Detrimental non-target exposure to ARs has been shown in numerous populations of
34 predators in Europe and North America (Christensen et al., 2012; López-Perea et al., 2015; Riley
35 et al., 2007; Shore et al., 2003; Thomas et al., 2017). These effects can be significant, with
36 population-level effects from non-target exposure documented for mammals (Jacquot et al.,
37 2013) and raptors (Thomas et al., 2011). It is thought that species that regularly prey upon small
38 rodents are at higher risk of poisoning, due to the likelihood of consuming AR targeted species
39 (Hindmarch and Elliott, 2018). However, the primary consumption of AR baits by non-target
40 species, as well as the potential for SGARs to move through trophic levels, may lead to wider
41 contamination of terrestrial food chains (Hindmarch and Elliott, 2018; Thomas et al., 2011). If
42 such broadscale contamination is apparent, species at higher trophic levels may be at increased
43 risk of AR exposure (Riley et al., 2007; Thomas et al., 2011).

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44 It is necessary to understand the drivers of patterns in non-target AR exposure in order to
45 assess long-term effects and to develop effective mitigation. There are documented differences in
46 AR exposure of predators between the sexes (McDonald et al., 1998) and among age groups
47 (Christensen et al., 2012; Ruiz-Suárez et al., 2016). That said, local anthropogenic factors are
48 likely the most significant drivers of overall risk of non-target exposure. For example, human
49 population density and developed surface area have been linked to the probability and level of
50 AR exposure of numerous predators (Lohr, 2018; Lopez-Perea and Mateo, 2018; Nogueira et al.,
51 2015; Serieys et al., 2015). Agricultural AR use has also been suggested as the cause of non-
52 target poisoning of predators (Birks, 1998; Fourel et al., 2018; Hindmarch et al., 2017; Hughes et
53 al., 2013), but only a few recent studies have found empirical evidence of this relationship
54 (Coourdassier et al., 2019; López-Perea et al., 2018; Rial-Berriel et al., 2021; Sainsbury et al.,
55 2018).

56 AR use is largely unmonitored in Australia and recent work has highlighted the need for
57 the evaluation of its effects on Australasian taxa (Lohr, 2018; Lohr and Davis, 2018). The
58 Tasmanian wedge-tailed eagle (*Aquila audax fleayi*) is a subspecies of wedge-tailed eagle
59 endemic to the Australian island of Tasmania (Commonwealth of Australia, 1999). With the loss
60 of the thylacine (*Thylacinus cynocephalus*) and recent declines in populations of Tasmanian
61 devils (*Sarcophilus harrisii*), the wedge-tailed eagle serves a particularly important ecological
62 function as one of the few remaining top predators in Tasmanian ecosystems. The subspecies is
63 listed as endangered (Commonwealth of Australia, 1999; State Government of Tasmania, 1995),
64 with conservation concern based upon a series of threats, including nest failures caused by
65 anthropogenic disturbance, low breeding success rates, habitat loss, collisions with

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66 anthropogenic structures, lead poisoning, and illegal persecution (Bell and Mooney, 1998;
67 Mooney and Holdsworth, 1991; Pay et al., 2020; Threatened Species Section, 2006).

68 ARs are not recognized as a significant threat to the Tasmanian wedge-tailed eagle
69 population, as the species generally avoids areas of high human population density, and rodents
70 represent a very small portion of their diet (Marchant and Higgins, 1993). That said, wedge-
71 tailed eagles show a high sensitivity to pindone (Martin et al., 1994); this AR is used to control
72 European rabbit (*Oryctolagus cuniculus*) populations, a primary prey species of wedge-tailed
73 eagles (Debus et al., 2007; Marchant and Higgins, 1993). Furthermore, if ARs are moving
74 through Tasmania's food chains, then the high trophic position of the wedge-tailed eagle may
75 increase their susceptibility to exposure to various AR compounds. Finally, because of the long-
76 lived and slow breeding life history strategy of this species, it is likely highly vulnerable to
77 increased mortality rates brought on by toxicants such as ARs.

78 Our aims in this research were to determine to what extent Tasmanian wedge-tailed
79 eagles are exposed to ARs, and to investigate the factors that influence AR exposure in the
80 population. Specifically, we evaluated (1) liver tissue concentrations of individual ARs known to
81 be used in Tasmania and the total SGAR concentration of each individual eagle; and the
82 relationship between both (2) total liver SGAR concentration and (3) probability of exposure
83 with intrinsic (age and sex) and extrinsic (human population density, agricultural land use, and
84 year of death) factors.

85 **2. Methods**

86 *2.1. Study area*

87 This study was conducted on mainland Tasmania, an island state located 240 km south of
88 continental Australia. Tasmania covers an area of 68,150 km², with an estimated human
89 population of 520,830 (Australian Bureau of Statistics, 2018; Figure 1b). Areas of minimal land
90 use, nature conservation and other protected areas account for 50.3% (34,280 km²) of the
91 Tasmanian land area (DPIPWE, 2015). Agriculture occupies 18,900 km² (27.7%; DPIPWE,
92 2015) mostly in the north and east of the state (Figure 1c). The Tasmanian agricultural land area
93 is comprised of modified pastures (75.4%), native vegetation pastures (14.5%), irrigated crops
94 (8.8%), and non-irrigated crops (1%; DPIPWE, 2015).

95

96 *2.2. Sample collection*

97 Eagles were collected as carcasses found opportunistically throughout Tasmania (Figure
98 1a) between 1996 and 2018, by government departments, various industries, and volunteers. All
99 carcasses were placed in -20°C freezer storage by the Department of Primary Industries, Parks,
100 Water and the Environment (DPIPWE, Threatened Species Section, Hobart, Tasmania) and the
101 Tasmanian Museum and Art Gallery (TMAG, Collection and Research Facility, Rosny,
102 Tasmania). Data recorded for each carcass included location and the date the carcass was found.
103 We thawed the carcasses and harvested tissues from them between May 2017 and March 2018.
104 We collected a whole liver lobe and a muscle sample from each carcass. The tissue samples were
105 stored at -20°C until sample preparation, when we thawed them at room temperature. We
106 weighed out a 4 g (\pm 0.1 g) wet weight sample from the middle of each liver lobe using a digital

107 balance (precision ± 0.0001 g (Mettler Toledo, US). New scalpel blades and gloves were used
108 between samples during collection and preparation to prevent cross contamination.

109

110 2.3. Residue analysis

111 2.3.1. Sample preparation

112 All toxicological analyses were carried out at Edith Cowan University Analytical Facility
113 (Joondalup, Western Australia). Each liver sample was freeze-dried and homogenized.
114 Homogenized samples were transferred into centrifuge plastic tubes (15 ml) and 10 ml of
115 acetonitrile was added to the tubes with a 10 μ l (10 ng/ μ l) solution containing deuterated
116 surrogates. Analytes were extracted using a sonication bath (15 minutes sonication for each
117 aliquot). After extraction, samples were centrifuged at 3,247 relative centrifugal force (rcf) for 5
118 minutes, transferred to a new centrifuge tube with 2 ml of hexane, vortexed for 5 minutes and
119 centrifuged at 3,247 rcf for a further 5 minutes. Each sample was then evaporated and
120 reconstituted in 400 μ l of 50:50 ACN/H₂O solution. The final extracts were transferred to 2 ml
121 Teflon-lined vials and stored at 0–4°C until analysis.

122 2.3.2. LC-MS analysis

123 Liver samples were analyzed for ARs registered for use in Australia (Australian
124 Pesticides and Veterinary Medicines Authority, 2019). Concentrations of five SGARs
125 (brodifacoum, bromadiolone, difethialone, difenacoum and flocoumafen) and three FGARs
126 (coumatetralyl, pindone and warfarin) (see Appendix Table A.1 for the manufacturers of the
127 analytical standards and surrogates) were evaluated using a TSQ Quantiva triple quadrupole
128 Mass Spectrometer (LC-MS) from Thermo Fisher (Thermo Fisher Scientific Corporation, US)
129 (see Appendix B for details of the chromatographic method). Calibration curves and recovery

130 rates for each analytical run were calculated using organic chicken livers spiked with three
131 working solutions of each analytical standard. Recovery rates for the target ARs averaged 96.75
132 %, whilst limits of detection (LOD) and limits of quantification (LOQ) ranged from 0.0005–
133 0.0125 mg/kg and 0.001–0.025 mg/kg respectively (Appendix Table A.2). All detections that
134 were > LOD but < LOQ were reported as present at trace levels. Three organic chicken liver
135 blanks were included in each run to monitor cross-contamination. Every 10th sample was
136 reinjected for a duplicate read (average percentage relative standard deviation of recoveries
137 (RSD) 4.1%) and duplicate blind sample extractions were carried out for five randomly selected
138 samples (average RSD 4.1%). Concentrations were reported on a dry weight basis (mg/kg dw).

139

140 *2.4. Potential drivers of AR exposure*

141 We evaluated potential drivers of AR exposure as a response to a suite of intrinsic and
142 extrinsic explanatory variables. The intrinsic variables we considered were the sex of the bird
143 (determined genetically using muscle tissue; Appendix C) and its age (broadly characterized into
144 adults and pre-adults based on plumage; Appendix D). Extrinsic explanatory variables were the
145 year in which the carcass was found, and both the mean human population density per km²
146 (Australian Bureau of Statistics, 2018) and the proportion of agricultural area (DPIPWE, 2015)
147 in the area surrounding where each carcass was found. Areas we categorized as agricultural
148 included all types of animal production (intensive animal production, native vegetation grazing,
149 and modified pastures grazing) and all types of horticulture (both non-irrigated, and irrigated
150 cropping; see Appendix E). We defined the area around where each carcass was found based on
151 the size of the estimated home range of adult and pre-adult eagles (25 km² for adults and 420
152 km² for pre-adults; see Appendix F). We buffered each carcass location by an area corresponding

153 to the age-specific home range and calculated the mean human population density per km² and
154 the proportion of agricultural land within the buffered area. To maximize accuracy in estimates
155 of spatial predictor variables, both human population density and agricultural land use area were
156 calculated from data as close to the year the carcass was found as possible (maximum differences
157 between year of death and spatial data were six years for human population and five years for
158 agricultural land use).

159

160 2.5. Data analysis

161 We performed all statistical analyses in R, version 3.2.0 (R Core Team, 2016). We
162 analyzed the data using censored data techniques (R packages NADA; Lee, 2017, and Survival;
163 Therneau, 2018) as some AR concentrations were below the LOD of the LC-MS.

164 2.5.1. Individual AR and total SGAR concentration

165 We used a Kaplan-Meier cumulative probability distribution (NADA function ‘cenfit’) to
166 calculate censored summary statistics (mean, median and standard error) of each AR compound
167 and the total SGAR concentration for each individual eagle. We also calculated analogous
168 standard (non-censored) summary statistics for only the eagles in which ARs were detected.
169 Creating these analogous summary statistics facilitated comparisons among our study and prior
170 work as other studies have used this approach (e.g. Hughes *et al.*, 2013; Lopez-Perea and Mateo,
171 2018). To facilitate comparisons to other studies, we calculated summary statistics on a wet
172 weight basis. To do this we converted dry weight concentrations (provided in mg/kg dw) to wet
173 weight (mg/kg ww) by multiplying the dry weight concentrations by the dry to wet weight ratios
174 of each sample.

175 We used total SGAR concentrations to estimate the effects of over-all SGAR
176 contamination due to their similar mode of action (Rattner and Harvey, 2021). FGARs were not
177 included in the summed concentrations due to large differences in molecular weight, potency and
178 half-life compared to SGARs (Rattner and Harvey, 2021). To estimate potential toxicological
179 effects of the total SGAR concentrations detected, we used published contamination thresholds
180 (see Lohr, 2018) as follows: (i) 0.001–0.01 mg/kg ww, probably no toxicity; (ii) 0.01–0.1 mg/kg
181 ww, unlikely lethal / possible toxicity; (iii) 0.1–0.5 mg/kg ww, possibly lethal / likely toxicity;
182 (iv) 0.5–0.7 ww, probably lethal; (v) > 0.7 mg/kg ww, lethal. We used the converted wet weight
183 concentrations for this evaluation as the thresholds were based on wet weight concentrations.

184 2.5.2. *Correlates of degree and likelihood of exposure*

185 We explored relationships between the extrinsic and intrinsic explanatory variables (age,
186 sex, year of death, human population density, and proportion of agricultural area) and total
187 SGAR concentration with left-censored regression models (Helsel, 2012; Survival function
188 ‘survreg’). We assigned censored data the corresponding LOD value with an indicator variable
189 denoting the observation as below the LOD. Uncensored data were assigned the total liver
190 SGAR concentration measured by the LC-MS and an indicator variable denoting the observation
191 as not censored. The correlation of predictor variables was checked before inclusion in the
192 models, and any correlated predictors (Pearson’s $r > 0.3$) were not included in the same model.
193 The dependent variable in these models was the total liver SGAR concentration (mg/kg dw) for
194 each sample. Our initial model set included all possible combinations of submodels. We used
195 corrected Akaike’s Information Criterion (AICc) to rank model performance. We excluded
196 models in the candidate set if they had an AICc value greater than six Δ AICc (Richards, 2005).
197 The use of AICc as the sole selection criterion may select overly complex models, thus we

198 considered only those models that had AICc values smaller than all the simpler models within
199 which they were nested (Richards, 2008). We based biological inferences on the coefficients of
200 the top-performing model and considered a parameter to have strong support if it was included in
201 all candidate models.

202 We also explored the relationship between the same suite of extrinsic and intrinsic
203 predictor variables with the probability of AR residues (both of SGARs and FGARs) being
204 detected using a binomial generalized linear model with logit link function. The dependent
205 variable in these models was whether the eagles were exposed (AR concentrations > LOD) or
206 unexposed (AR concentrations < LOD). We again considered all possible parameter
207 combinations and retained models in the candidate set that were both within six Δ AICc and had
208 AICc values smaller than all the simpler models within which they were nested.

209 **3. Results**

210 We analyzed tissue from 50 eagle carcasses that were collected between 1996 and 2018,
211 although most were collected after 2006 (n = 37). All birds were successfully sexed and aged,
212 with 41 eagles identified as pre-adult, and 22 as female. Data available for the sampled carcasses
213 included location (n = 50; Figure 1a) and year the carcass was found (n = 50).

214

215 *3.1. Individual AR and total SGAR concentration*

216 AR residues were detected in 74% of wedge-tailed eagles included in the study (Table 1).
217 Residues of more than one AR compound were detected in 38% of the birds, and 12% of birds
218 had three different compounds detected. The mean total SGAR concentrations of birds in which

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219 SGARs were detected was 0.143 mg/kg ww (\pm SE 0.031) and the censored mean of the entire
220 study sample was 0.100 mg/kg ww (\pm SE 0.023). The majority of AR residues were SGARs.
221 Brodifacoum (56% of birds), flocoumafen (40%) and bromadiolone (22%) were the most
222 predominant SGARs detected. FGARs were only detected in three individuals (one of these
223 individuals was also exposed to the SGAR flocoumafen). Warfarin was detected at very low
224 concentrations (< 0.01 mg/kg ww) in two birds and coumatetralyl was detected in one bird.

225 We recorded a potentially lethal total liver SGAR concentration (> 0.1 mg/kg ww;
226 Newton *et al.*, 1999) in 11 of the wedge-tailed eagles sampled (22%; 31% of those in which
227 SGARs were detected; Figure 2). Furthermore, concentrations were above probably lethal levels
228 of > 0.5 mg/kg ww in 4% of the eagles (6% of those in which SGARs were detected). That said,
229 liver AR concentrations do not allow the confirmation of lethality without a necropsy of the
230 animal identifying signs of toxicity.

231

232 3.2. Correlates of degree of exposure

233 The top-performing censored regression model suggested that total liver SGAR
234 concentration (mg/kg dw) was driven most strongly by the year the carcass was found, the
235 amount of agricultural area, and the human population density in the area around where the
236 carcass was found (see Appendix Table G.1). This model was 42.83 times more likely than the
237 null model. A simpler model that excluded human population density was also retained in the
238 candidate model set (Table 2). The year the carcass was found and agricultural area were
239 included in both candidate models, suggesting that these variables were the most important to
240 explaining total liver AR concentration. Coefficients of the best performing model indicated that
241 year of death, agricultural area, and human population density were all positively associated with

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242 total AR concentration (Table 3, Figure 3a). The model suggested that a 10% increase in
243 agricultural habitat in the area around where the bird died would result in an increase in liver AR
244 concentrations by a factor of 2.11. Likewise, each later year in the study was estimated to
245 increase AR concentrations by a factor of 1.40. The relationship between total AR concentration
246 and human population density suggested an increase in 100 inhabitants per km² would increase
247 total AR concentration by a factor of 7.23.

248

249 *3.3. Correlates of likelihood of exposure*

250 The top-performing binomial model to explain the probability of an eagle being exposed
251 to ARs included the year the carcass was found and the proportion of agricultural area within the
252 area around where the carcass was found (see Appendix Table G.2). The candidate model set
253 included two simpler models, including the null model (Table 2), although the top-performing
254 model was 8.9 times more likely than the null model based on AICc weight. Coefficients of the
255 top-performing model indicated that the probability of ARs being detected increased with
256 carcasses found more recently and in areas with higher proportions of agricultural area (Table 3).
257 The odds of ARs being detected in a carcass were 1.46 times greater for each 10% increase in
258 agricultural habitat proportion in the area around where the bird died and 1.21 times greater for
259 each advancing year of the study period (Figure 3b, Appendix Figure G.1).

260 **4. Discussion**

261 The frequency and magnitude of AR exposure in Tasmanian wedge-tailed eagles, and
262 their correlation to agricultural areas and human population density, have several implications
263 for our understanding of rodenticide exposure and the Tasmanian ecosystem. First, rodenticide

264 exposure is high among these birds, suggesting that rodenticides are frequently finding their way
265 into top predators in the ecosystem. Furthermore, extrinsic (i.e. agricultural area, human
266 population density, and year of death) rather than intrinsic factors (i.e. age, sex) influence the
267 probability of exposure to ARs and total SGAR concentration. These findings illustrate how AR
268 exposure of the Tasmanian wedge-tailed eagle is driven by anthropogenic processes and thus
269 identify directions to solve this conservation problem.

270

271 *4.1. Individual AR and total SGAR concentration*

272 The high prevalence of SGARs detected in our study is consistent with research
273 implicating SGARs as the predominant cause of non-target AR exposure of predators (Lohr,
274 2018; López-Perea et al., 2015). SGARs brodifacoum, bromadiolone, and flocoumafen
275 accounted for 99.6% of the total AR concentrations observed in the Tasmanian wedge-tailed
276 eagle. The first two of these are the AR compounds most commonly identified in non-target
277 predators in numerous ecosystems worldwide (Hosea, 2000; Koivisto et al., 2016; Langford et
278 al., 2013; Ruiz-Suárez et al., 2014; Sharp et al., 2005). The extent of the flocoumafen
279 contamination we detected is more surprising and represents one of the highest exposure rates
280 documented globally (see Appendix Table G.3). Flocoumafen is only available through
281 wholesale outlets in Tasmania, suggesting that agricultural asset protection and professional pest
282 controllers could be important sources of non-target AR exposure in Australia.

283 The low concentrations and frequency of detections of FGARs also corresponds with
284 findings for other species, both in Australia and globally (Cypher et al., 2014; Lohr, 2018;
285 Murray, 2020, 2017). This was of particular interest in the case of the FGAR pindone. We
286 expected pindone to be the AR most frequently detected in wedge-tailed eagles, since it targets a

287 common prey item for the species (the European rabbit). Our low detection of FGARs could be
288 due to their characteristics - their shorter half-life and lower toxicity - and, in the case of
289 pindone, its localized use in targeted control efforts (Lohr, 2018). Although this low rate of
290 detection may suggest FGARs pose a lower risk of non-target exposure, their characteristics may
291 also impede their detection in studies using opportunistic sampling and prolonged tissue storage
292 (Herring et al., 2017; Rattner et al., 2014), consequently underestimating their true prevalence in
293 the Australian environment.

294 The frequency and magnitude of AR exposure in the Tasmanian wedge-tailed eagle was
295 high for an *Aquila* species. Raptor studies showing comparable AR detection rates typically
296 involve smaller species known to be at risk due to their dietary specialization on rodents
297 (Christensen et al., 2012; López-Perea et al., 2015; Walker et al., 2011). The proportion of birds
298 we observed with concentrations > 0.2 mg/kg ww (16%) is substantially higher than that found
299 in congeners (0–6%; Hosea, 2000; Langford et al., 2013; Sánchez-Barbudo et al., 2012), and the
300 highest concentration of an SGAR we detected in an individual (0.635 mg/kg ww of
301 Brodifacoum) is substantially higher than the highest concentration of an AR previously reported
302 in an *Aquila* species (0.154 mg/kg ww of Bromadiolone; Langford et al., 2013). Both the
303 censored mean SGAR concentrations of all eagles sampled (0.100 mg/kg ww) and the mean only
304 of those with detected SGAR levels (0.143 mg/kg ww) were higher than mean concentrations
305 reported for congeners (0.006–0.073 mg/kg ww; Langford et al., 2013; Sánchez-Barbudo et al.,
306 2012), but lie within the range of values reported for other raptors exposed to SGARs (0.005–
307 0.413 mg/kg ww; Thomas et al. 2011; Christensen 2012; Lohr 2018).

308 The high exposure to SGARs in the Tasmanian wedge-tailed eagle, a species not known
309 to regularly prey upon synanthropic rodents, supports the suggestion that apex predators are

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310 vulnerable to SGARs (López-Perea et al., 2015; Riley et al., 2007). The long half-life and
311 persistence of SGARs gives these compounds the capacity to move through food chains (López-
312 Perea et al., 2015), a theory evidenced by the presence of ARs in apex predators (Riley et al.,
313 2007). The wedge-tailed eagle preys upon several carnivorous species that are known to
314 consume synanthropic rodents. For example predatory and scavenging species such as forest
315 ravens (*Corvus tasmanicus*), kookaburras (*Dacelo novaeguineae*), common brush-tail possums
316 (*Trichosurus vulpecula*), cats (*Felis catus*), and other raptors have been recorded in wedge-tailed
317 eagle diets (Marchant and Higgins, 1993). The potential for SGARs to move through multiple
318 trophic levels may therefore be causing extensive contamination of Tasmania's terrestrial food
319 chains (Thomas et al., 2011). If this is the case, then numerous other predatory species may be at
320 risk in the region, including the endangered Tasmanian devil and eastern quoll (*Dasyurus*
321 *viverrinus*; IUCN, 2020).

322 The high exposure we detected may also be driven by the improper use of ARs and non-
323 target AR vectors. The use of SGARs in Australia does not require a license, products can be
324 easily purchased in large quantities, and awareness of use guidelines may be low. If SGARs are
325 not being used as directed, numerous non-rodent species may consume the poisons and act as AR
326 vectors to predators. Furthermore, ARs have recently been detected in Australian reptile species;
327 this exposure could be through direct consumption of ARs used correctly, since these species are
328 small enough to enter AR bait boxes (Lettoof et al., 2020). Reptiles are prey for wedge-tailed
329 eagles, as well as other predators, and may therefore have a role as AR vectors in Australia.

330

331 *4.2. Correlates of AR exposure*

332 The positive association between hepatic AR concentrations and human population
333 density and agricultural land use may indicate localized use that is having wider scale effects.
334 AR residues in predators have been linked to human population density (López-Perea et al.,
335 2018, 2015), the amount of urbanized area (Coeurdassier et al., 2019; Lohr, 2018; Serieys et al.,
336 2015), and the amount of both arable and pastoral agriculture (Coeurdassier et al., 2019; López-
337 Perea et al., 2018; Sainsbury et al., 2018). These relationships are unsurprising in study species
338 known to use urban and agricultural habitats. However, Tasmanian wedge-tailed eagles are less
339 associated with densely populated areas. Although human population growth has been relatively
340 low in Tasmania for the past two decades, there has been an increase in the number of residences
341 built in more rural and natural areas and agricultural development has expanded (Australian
342 Bureau of Statistics, 2018). Such practices may introduce ARs into more natural areas.
343 Furthermore, if ARs are passing through multiple trophic levels, they will spread more widely
344 from the initial bait. The effects of these more remote developments and agricultural activities
345 may therefore have incommensurately greater effects on predatory species than suggested by the
346 landscape footprint.

347

348 *4.3. Recent increases in AR exposure*

349 The higher total SGAR concentrations and probability of AR exposure of the birds that
350 had died more recently could be due to either the increased exposure to ARs over time or the
351 degradation of the compounds with prolonged storage. Although SGAR residues are stable
352 within tissues over the short-term (Galocchio et al., 2014; Jin and Chen, 2006), the effects of
353 long-term -20°C freezer storage on tissue residues is not well understood, with studies

354 documenting various rates of degradation (e.g. 6–41% over 0.5–3 years; P. Fisher unpublished
355 data; Vindenes *et al.*, 2008). Despite this, patterns in the AR concentrations we detected are
356 consistent with increased probability of exposure over time. There would need to be a substantial
357 reduction in AR residues (much greater than the degradation rates documented) for an AR-
358 exposed bird to be considered unexposed in our study, as the lowest AR concentration we
359 detected was still 200% greater than the associated LOD. Consequently, the increased AR
360 concentrations in Tasmanian wedge-tailed eagles that had died more recently is more likely due
361 to increases or changes in AR use in Tasmania than to sample degradation. However, there is no
362 information available on the volume of ARs used in Australia (Lohr and Davis, 2018), which
363 impedes our quantification of the relationship between AR application and non-target AR
364 exposure.

365

366 *4.4. Conservation implications*

367 Our results indicate that AR exposure is likely a significant factor to consider in the
368 conservation management for the Tasmanian wedge-tailed eagle. This is true even given the
369 potential biases inherent to the non-random carcass collection we relied on to gather samples.
370 AR studies using opportunistic samples may inflate the proportion of animals with sub-lethal AR
371 concentrations detected and underestimate the proportion of birds detected with fatal levels
372 (Lohr, 2018; Newton *et al.*, 1990). We found exposure at rates that are high compared to other
373 studies using similar sampling methods.

374 The use of AR concentration thresholds to interpret the likely physiological result has
375 limitations due to inter- and intra-specific variation in susceptibility to toxicity (Rattner and
376 Harvey, 2021 ; Thomas *et al.*, 2011). That said, concentrations in 22% of the birds we studied

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377 were well above the potentially lethal range reported for European barn owls (*Tyto alba*; > 0.1
378 mg/kg; Newton et al. 1999). Furthermore, 56% had levels that can cause symptoms of toxicity
379 (> 0.01 mg/kg; (Lohr, 2018; Murray, 2018);. These comparative data therefore suggest that the
380 level of exposure we detected indicates that AR exposure could be influential to survival and
381 possibly conservation of these birds.

382 Our findings underscore the importance of efforts to address non-target AR exposure in
383 Australian wildlife. SGARs are currently registered for domestic (non-professional) use in
384 Australia (Australian Pesticides and Veterinary Medicines Authority, 2019), despite increasing
385 regulation and monitoring in other countries (USEPA, 2008). Increased legislative control of
386 SGARs and removal from public retail have been suggested as steps to reduce the ecological
387 effects of SGAR use in Australia (Lohr and Davis, 2018). However, our findings of an
388 association between agriculture and AR concentrations in the Tasmanian wedge-tailed eagle, as
389 well as widespread contamination of an AR not readily available for residential use
390 (flocoumafen), suggests that professional pest control may also be an important cause of non-
391 target AR exposure. Addressing mechanisms of spread from both professional and non-
392 professional application of SGARs may therefore be important to reducing AR exposure in
393 Tasmanian wedge-tailed eagles and other Australian wildlife.

394

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AR exposure in an Australian top predator

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- 602

603

604 Table 1. Summary statistics describing liver AR concentrations of each AR assessed and total liver SGAR concentration of Tasmanian
 605 wedge-tailed eagle carcasses collected between 1996 and 2018. Non-censored and censored summary statistics are presented. Non-
 606 censored statistics were calculated using only the eagles with detected AR concentrations. Censored summary statistics consider all
 607 individuals and account for unknown values below the corresponding limit of quantification (LOQ). All summary statistics are
 608 reported on a wet weight basis.

	Brodifacoum	Bromadiolone	Coumatetralyl	Difenacoum	Difethialone	Flocoumafen	Pindone	Warfarin	Total SGAR
Not censored									
LOQ (mg/kg)	0.005	0.001	0.002	0.0025	0.010	0.0025	0.025	0.002	NA
Birds exposed (%)	28/50 (56%)	11/50 (22%)	1/50 (2%)	0/50 (0%)	0/50 (0%)	20/50 (40%)	0/50 (0%)	2/50 (4%)	37/50 (74%)
Max (mg/kg ww)	0.635	0.241	0.014	0.000	0.000	0.348	0.000	0.002	0.651
Min (mg/kg ww)	0.003 ^a	0.003	0.014 ^a	0.000	0.000	0.002 ^a	0.000	0.001 ^a	0.002
Mean (mg/kg ww)	0.136	0.045	0.014	0.000	0.000	0.035	0.000	0.002	0.143
Median (mg/kg ww)	0.072	0.023	0.014	0.000	0.000	0.003	0.000	0.002	0.074
SE (mg/kg ww)	0.030	0.021	0.000	0.000	0.000	0.022	0.000	0.001	0.031
Censored									
Mean (mg/kg ww)	0.077	0.012	0.014	NA	NA	0.015	NA	0.002	0.100
Median (mg/kg ww)	0.011	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	0.017
SE (mg/kg ww)	0.019	0.005	NA	NA	NA	0.009	NA	0.000	0.023

609 ^aTrace value

610

AR exposure in an Australian top predator

611 Table 2. Candidate models relating total liver SGAR concentrations and probability of ARs being detected in Tasmanian wedge-tailed
 612 eagle carcasses collected between 1996 and 2018 to intrinsic and extrinsic factors^a.

Rank	Model variables	df	AICc	AICc weight
Total liver AR concentration				
1	Year of death (+); Agricultural area (+); Human population density (+)	5	93.170	0.795
2	Year of death (+); Agricultural area (+)	4	95.885	0.205
Probability of exposure				
1	Year of death (+); Agricultural area (+)	3	55.010	0.712
2	Year of death (+)	2	57.471	0.208
3	<i>Null model</i>	1	59.389	0.079

613 ^a Only models that were both within six Δ AICc and had AICc values smaller than all the simpler models within which they were nested were
 614 retained in in the candidate model set (Richards, 2008). For year of death, “+” indicates association to carcasses found more recently.

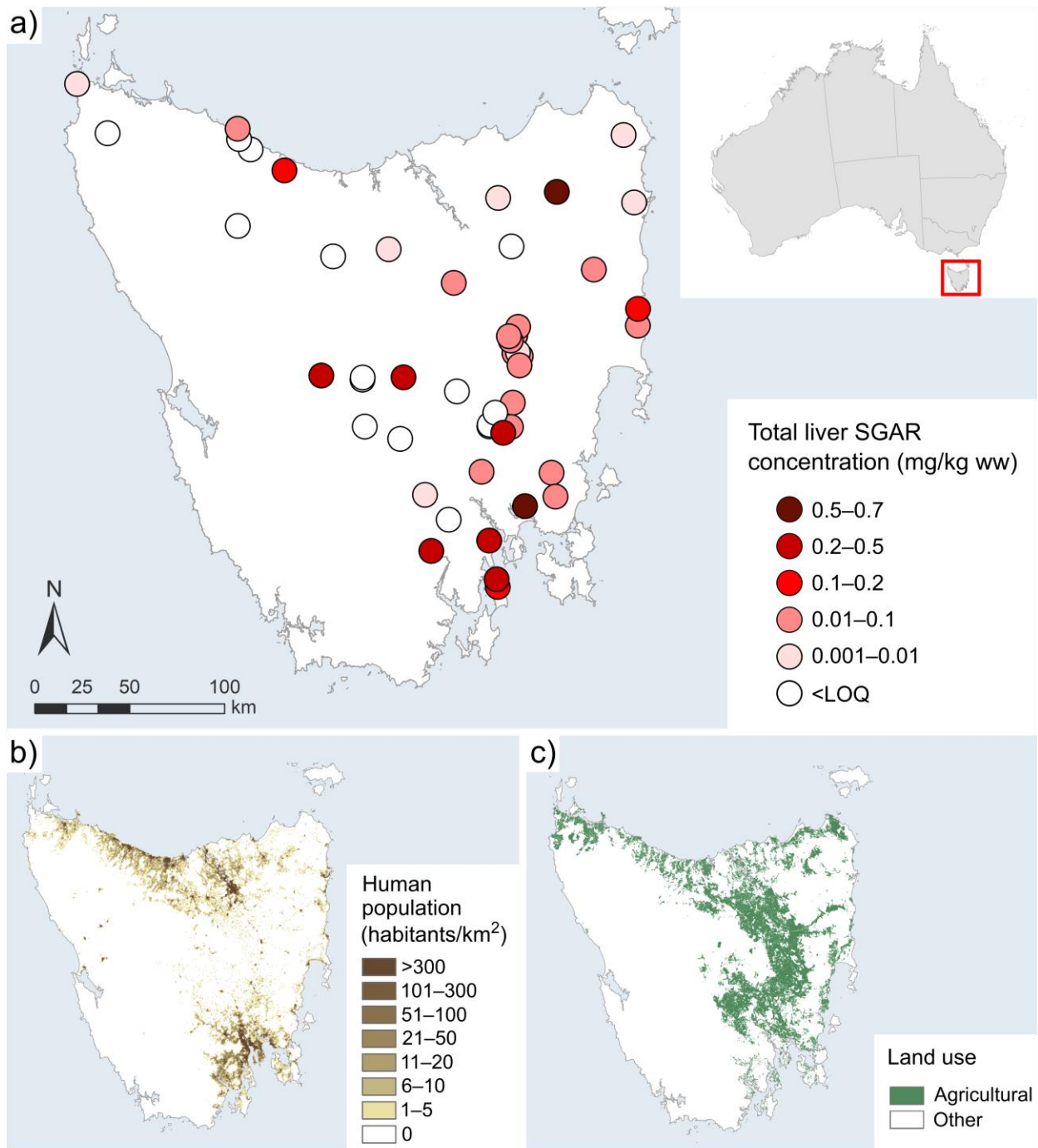
615

616 Table 3. Model coefficients for top performing models describing the estimated effect of each
 617 variable on total liver SGAR concentrations (censored regression) and the probability of an eagle
 618 being exposed to ARs (binomial probability).

Model	Parameter	Estimate	95% CI		z
			Lower	Upper	
Total liver AR concentration	Intercept	-12.105	-16.669	-7.542	-5.20
	Year of death	0.337	0.137	0.536	3.31
	Agricultural area	0.749	0.248	1.249	2.93
	Human population density	1.978	0.306	3.650	2.32
Probability of exposure	Intercept	-2.945	-6.313	-0.165	-1.93
	Year of death	0.193	0.053	0.365	2.48
	Agricultural area	0.375	0.035	0.781	2.02

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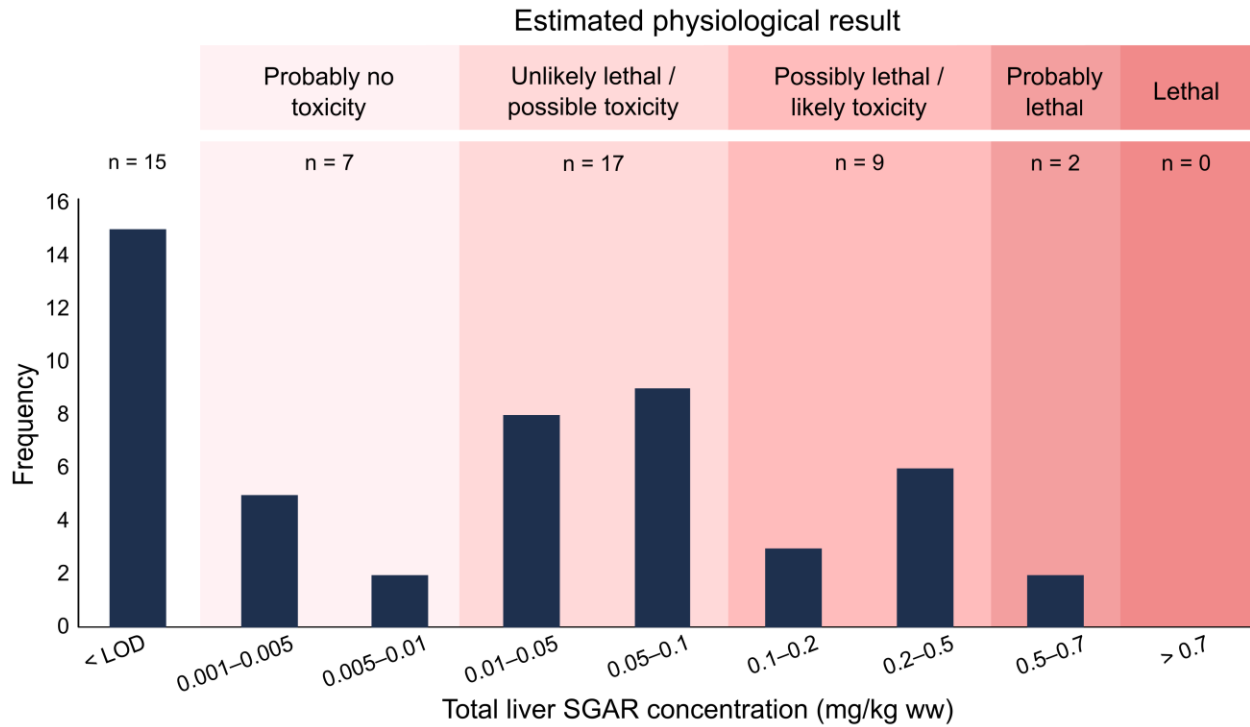


621

622 Figure 1. a) Location and liver total SGAR concentration threshold for 50 Tasmanian wedge-
 623 tailed eagle carcasses collected between 1996 and 2018. Maps b) and c) indicate the spatial
 624 distribution of the Tasmanian human population (2016 data; Australian Bureau of Statistics,
 625 2018) and agricultural land use area (2015 data; DPIPWE, 2015) respectively.

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628

629 Figure 2. Total liver SGAR concentrations (mg/kg ww) for Tasmanian wedge-tailed eagles that
 630 died between 1996 and 2018 (n = 50). The number of eagle carcasses with liver SGAR
 631 concentrations within each toxicity threshold proposed by Lohr (2018) is presented.

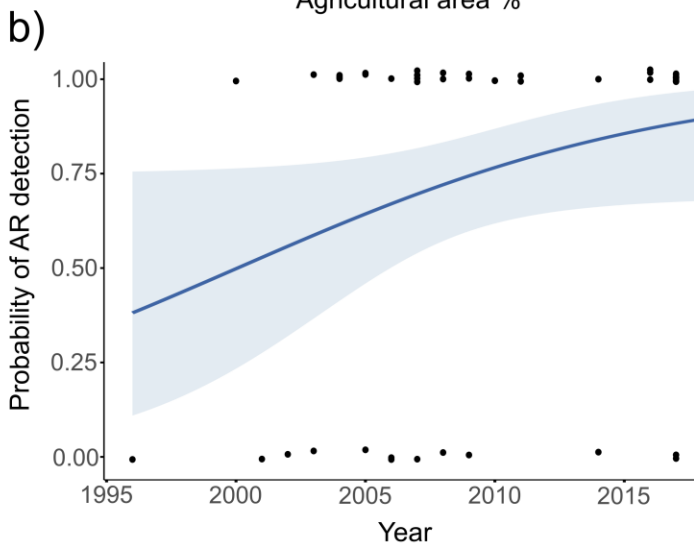
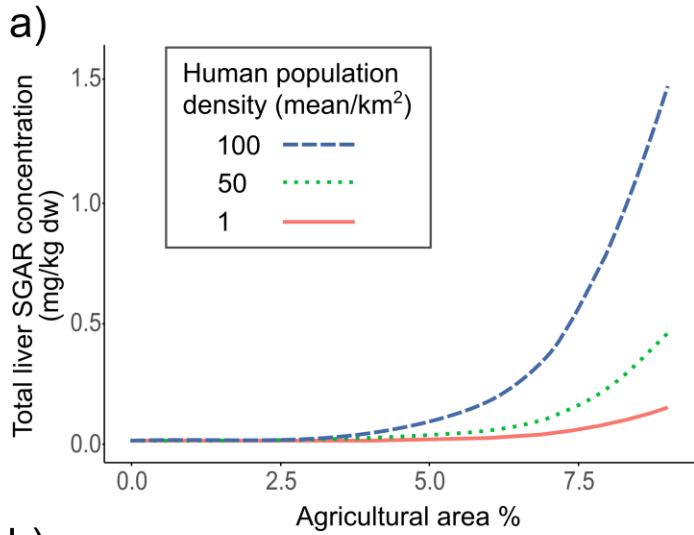
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638 Figure 3. a) Predicted response of total liver SGAR concentrations (mg/kg dw) in Tasmanian
 639 wedge-tailed eagle carcasses as a function of the proportion of agricultural land area and mean
 640 human population density in the area around where the bird died. The three lines are the
 641 estimated response of liver AR concentration with human population per km² held at three
 642 levels. Year of carcass discovery is held at its mean. b) Logistic plot of the effect of year of death
 643 on the probability of AR exposure. Agricultural land area is held at its mean.

644