
**TEMPORAL AND SPATIAL VARIABILITY OF
THE LIFE HISTORY CHARACTERISTICS OF
SAND FLATHEAD, *PLATYCEPHALUS*
BASSENSIS.**

ALI BANI, BSc, (MSc) TEHRAN UNIVERSITY

**SUBMITTED IN FULFILMENT OF THE REQUIREMENTS FOR
THE DEGREE OF DOCTOR OF PHILOSOPHY**

UNIVERSITY OF TASMANIA

OCTOBER 2005

DECLARATION OF ORIGINALITY

This thesis contains no material which has been accepted for a degree or diploma by the University or any other institution, except by way of background information and duly acknowledged in the thesis, and to the best of my knowledge and belief no material previously published or written by another person except where due acknowledgement is made in the text of the thesis.

Ali Bani



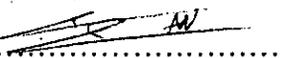
Date

20/10/05

STATEMENT OF ACCESS

This thesis may be made available for loan. Copying any part of this thesis is prohibited for two years from the date this statement was signed; after that limited copying is permitted in accordance with the *Copyright Act 1968*.

Ali Bani



Date

20/10/05

ABSTRACT

Spatial and temporal variability in life history characteristics of sand flathead *Platycephalus bassensis*, were explored in the context of energy tradeoffs between growth and reproduction. Given the similarity in environmental conditions and somatic condition of individuals and the populations, this study was able to examine whether and to what extent spatial variability in dynamics of recreationally fished populations was associated with variation in life history characteristics. This was achieved through intensive sampling at three locations (Georges Bay, Coles Bay, and the Tamar River) describing variability in demographic parameters, variability in reproductive life history characteristics, and the patterns of energy allocation to growth and reproduction.

Demography of sand flathead populations varied significantly among the three locations around the coastal waters of Tasmania. The proportion of older and larger fish was greater in Coles Bay, compared with the Georges Bay and the Tamar River populations. The absence of old (> 4 years) individuals in the Tamar Estuary was attributed to the movement of older fish out of the estuary. In contrast, less older and larger individuals in Georges Bay was hypothesised to be a function of greater mortality rates of fast-growing individuals. Similarity in growth rates of all examined populations during the first two years of life, before they suffer fishing pressure, suggests that environmental conditions at least for younger fish, were similar in all three locations.

Reproductive potential and spawning strategy of *P. bassensis* populations also differed spatially. Very low attainment of maturity in the Tamar River population during the spawning season, together with minimal gonad weight and lack of seasonal changes in gonad development, as well as the absence of older fish suggests

that the Tamar Estuary is most probably not part of the spawning grounds for *P. bassensis*. Duration of spawning activity in the Coles Bay population (October-March) was longer than for the Georges Bay population (September-November), suggesting a greater chance to spawn more batches of eggs during the spawning period in Coles Bay. Additionally, the Coles Bay population showed greater reproductive investment during the spawning season, compared to Georges Bay. As a result, the Coles Bay population, which consisted of relatively more larger individuals was expected to have a greater reproductive output compared with Georges Bay, which had predominantly smaller individuals.

For both mature and immature individuals differences in somatic and liver condition among the populations were relative small compared to the differences in life history parameters of growth and reproduction. Changes in liver lipid and water content of Coles Bay and Georges Bay individuals with reproductive state indicated that sand flathead used liver lipid for reproduction. No changes in muscle water content of mature individuals suggested that muscle is not a primary energy source for reproduction.

A comparison of the patterns of energy allocation and the condition of individuals from different populations indicated that variability in the life history traits can not be explained in the context of energy allocation. Absence of spatial variability in somatic condition and energy allocation suggests that significant spatial variability in *P. bassensis* life history traits may be a result of population responses to exogenous factors, e.g. fishing. Such a response can override other contributing factors in the life history variability and consequently drive population structure and dynamics.

ACKNOWLEDGMENTS

Assistance came from many directions during the course of this study. First, my special and sincere thanks to my supervisor Dr. Natalie Moltschaniwskyj for her continued support, guidance and encouragement throughout all phases of this study. Your supervision and help were sincerely appreciated, and made this study possible. Special thanks must also go to my co-supervisor Dr. Alan Jordan for providing an often-needed sounding board and constructive advice throughout the duration of this project.

Needless to say this study can never be done without the technical support of technicians of the aquatic centre of School of Aquaculture. I am indebted to them for their generous assistance in sampling. In particular I would like to acknowledge Greg Kent and Matt Foale for their friendly support. Thanks also to Natalie Moltschaniwskyj for her substantial assistance and contribution in the field. Thank you to all volunteers for their contribution in fishing; Don Ward, Tom Smith, Tiffany Wardley, Tim Green, Michael Steer, Ross Daghish, Mark Powell, and Shane Roberts.

I am very grateful to Prof. Ned Pankhurst for helping me to stage gonads, and providing me with invaluable direction in fish hormone treatment. Thanks also to Dr. Mark Porter for showing me hormone injection and for his assistance.

I am very appreciative to Prof. Chris Carter guidance for lipid analysis and for letting me use the facilities of the nutrition laboratory.

At last, but not least, I would especially like to thank my wife, Nasrin, for being my pillar of strength throughout the entire PhD experience. Without her support I would not have lasted the distance. This thesis is dedicated to you.

TABALE OF CONTENTS

Temporal and Spatial Variability of the Life History Characteristics of Sand Flathead, <i>Platycephalus bassensis</i>.	I
DECLARATION OF ORIGINALITY	II
STATEMENT OF ACCESS	II
ABSTRACT	III
ACKNOWLEDGMENTS	V
TABALE OF CONTENTS	VI
FIGURE LEGEND	VII
TABLE LEGEND	XI
Chapter One: General Introduction	1
1.1. POPULATION STRUCTURE	2
1.2. GENERAL OBJECTIVES	8
1.3. CHAPTER SCOPES	8
Chapter Two: Spatial and temporal variation in age, growth, and mortality of sand flathead, <i>Platycephalus bassensis</i>.	11
2.1. INTRODUCTION	12
2.2. MATERIALS AND METHODS	15
2.2.1. Study sites and sampling	15
2.2.2. Age determination	17
2.2.3. Analysis	19
2.3. RESULTS	21
2.3.1. Environmental conditions	21
2.3.2. Precision of age estimates and otolith examination	23
2.3.3. Size and age composition	24
2.3.4. Growth	31
2.3.5. Longevity and mortality	37
2.4. DISCUSSION	39
Chapter Three: Reproductive biology of sand flathead, <i>Platycephalus bassensis</i>.	47
3.1. INTRODUCTION	48
3.2. MATERIALS AND METHODS	52
3.2.1. Field sampling	52
3.2.2. Laboratory processing and histological analysis	54
3.2.3. Spawning frequency and hormone treatment	59
3.3. RESULTS	62
3.3.1. Gonad development	62
3.3.2. Reproduction and spawning strategy	67
3.3.3. Spawning periodicity	73
3.4. DISCUSSION	77
Chapter Four: Spatial and temporal variability in reproductive ecology of sand flathead, <i>Platycephalus bassensis</i>.	83
4.1. INTRODUCTION	84
4.2. MATERIALS AND METHODS	87
4.2.1. Study sites and sampling	87
4.2.2. Laboratory processing	87
4.2.3. Size and age at maturity	88
4.2.4. Analysis	88
4.3. RESULTS	89
4.3.1. Changes in gonad development	89
4.3.2. Gonad weight vs. size/age	95

4.3.3. Size and age at sexual maturity.....	95
4.3.4. Sex ratios.....	99
4.4. DISCUSSION.....	99
Chapter Five: Spatial variation in condition and patterns of repro-somatic investment in sand flathead, <i>Platycephalus bassensis</i>.....	106
5.1. INTRODUCTION.....	107
5.2. MATERIALS AND METHODS.....	110
5.2.1. Study sites and sampling.....	110
5.2.2. Laboratory processing.....	111
5.2.3. Analysis.....	112
5.3. RESULTS.....	114
5.3.1. Condition.....	114
5.3.2. Proximal analysis.....	119
5.4. DISCUSSION.....	125
Chapter Six: Synthesis and Further Directions.....	131
6.1. SUMMARY.....	132
6.2. SYNTHESIS.....	132
6.3. DIRECTIONS FOR FURTHER RESEARCH.....	137
References.....	140

FIGURE LEGEND

- Figure 1.1.** Schematic of population formation and contributing parameters in this process.....3
- Figure 2.1.** Map showing sampling locations for sand flathead.....16
- Figure 2.2.** The average of monthly water temperature and salinity near the benthos for each location.....22
- Figure 2.3.** The frequency distribution of the differences in the age of *P. bassensis* estimated using thin-section and burn and crack techniques, n=100.....23
- Figure 2.4.** Length-frequency distribution of *P. bassensis* in three locations. Arrows indicate the direction in which the observed frequencies differed from expected frequencies, generated under the assumption that size class was independent of location.....25
- Figure 2.5.** Length frequency distribution of *P. bassensis* at each location for 2001/02 and 2002/03 sampling years. Arrows indicate the direction in which the observed frequencies differed from expected frequencies generated, under the assumption that size class was independent of year of sampling.....26
- Figure 2.6.** Age frequency distributions of *P. bassensis* at three locations. Arrows indicate the direction in which the observed frequencies differed from expected frequencies, generated under the assumption that age class was independent of location.....28
- Figure 2.7.** Age frequency distribution of *P. bassensis* at each location for 2001/02 and 2002/03 sampling years. Arrows indicate the direction in which the observed frequencies differed from expected frequencies generated, under the assumption that age class was independent of year of sampling.....29
- Figure 2.8.** The mean age (a) and length (b) of *Platycephalus bassensis* for each location: Means with different letters above the bars represent groups that are statistically different. Values above letters represent the sample size.....31
- Figure 2.9.** The relationship between age and total length of *Platycephalus bassensis* in Georges Bay and the Tamar River at each sampling year. Values in parentheses are standard errors for the slope and intercept.....32
- Figure 2.10.** Von Bertalanffy growth curves for Coles Bay *Platycephalus bassensis* for each year of sampling and each sex. See Table 2.2 for estimates parameters for the four growth curves.....34
- Figure 2.11.** Approximate 95% confidence ellipses of the parameter estimates K (growth coefficient) and L infinity (mean asymptotic total length) of Von Bertalanffy growth curve for sand flathead caught from Coles Bay (following Kimura 1980).....35
- Figure 2.12.** (a) Comparison of slope of linear age-length relationship (+/- 95% Confidence Limit) and, (b) linear fit of this relationship among locations for *P. bassensis* < 5 years old. Coles Bay (solid line), Tamar River (dotted line), and Georges Bay (broken line). Asymmetric CLs resulted from back transformation of logarithmic values.....37
- Figure 2.13.** Age-based catch curve and estimates of *P. bassensis* mortality in the two populations. Mortality estimates per each year are; Coles Bay, 0.28 and Georges Bay, 0.59. Values in parentheses are standard errors for the slope and intercept.....38
- Figure 3.1.** Histological section of ovary showing oocytes development (stage 2-6). (A) Stage 2: A dark, dense and thick cytoplasm (cy) around a light nucleus (nu) of pre-vitellogenic oocytes (po); nucleoli (n) at periphery of nucleus. (B) Stage 3: Appearance of yolk vesicles (yv) in cytoplasm of primary vitellogenic oocyte (pvo). (C) Stage 4: Marked increase in secondary vitellogenic oocyte

(svo) size. Cytoplasm bounded by the zona radiata (zr), filled with yolk granules (yg) and oil vesicles. (D) Stage 5: Two nucleus (nu) migrated oocytes. (E) Stage 6: Onset of hydration (h), yolk granules coalesced to form yolk plates (yp). (F) Post-spawning: Folded postovulatory follicles (POF). Scale bars 0.1 mm.....63

Figure 3.2. Oocyte size frequency distribution in sand flathead at different stages of gonadal maturation. Due to the lack of more than 50% of oocyte types in stages 2 and 3, frequency of oocyte could not be statistically compared among stages.....64

Figure 3.3. Photomicrograph of whole oocytes of sand flathead representing unyolked (U), partially yolked (PY), yolked (Y), nucleus migrated (NM), and hydrated (H) oocytes. Translucent strip of perivitelline (arrows) in yolked oocyte changed, to a transparent cap in nucleus migrated oocyte. Scale bar 0.5 mm.....65

Figure 3.4. Frequency of different gamete stages in histological sections from each macroscopic testicular stage of sand flathead. The arrows indicate the direction in which the observed frequencies differed from expected frequencies, generated under the assumption that gamete type is independent of testis stage. n = number of individuals, with 500 observation per testis of each individual.....66

Figure 3.5. Photomicrographs of (A) spermatogenic and (B) spermiated testes from sand flathead, showing spermatogonia (SPG), primary and secondary spermatocytes (SC1 and SC2), spermatids (SPD), and spermatozoa (SPZ). Scale bars 100 μ m.....67

Figure 3.6. States of atresia in sand flathead ovaries described in Table 3.4. (A) State 1: A small proportions of yolked oocytes are affected by atresia. An alpha (α) stage of atretic oocyte beside two intact vitellogenic oocytes. (B) State 2: Predominance of atretic oocytes, here beta (β) stage, in ovary. (C) State 3: α , β , and gamma (γ) stages of atresia influenced in all oocytes. Scale bars 0.1 mm.....71

Figure 3.7. Deterioration of sand flathead POF following release of eggs. (A) Newly (< 6 h post-spawning) collapsed follicle with distinct granulosa (g) and theca (t) cells. Changes of POF structure at 24 h (B), 48 h (C), and 72 h (D) post-spawning. l=lumen. Scale bars 0.05 mm.....74

Figure 3.8. Proportion of mature female *Platycephalus bassensis* with early (structure similar to 24 h age POF or younger) or late-stage (structure similar to 36 h age POF or older) POF at different moon phase in October and December. Values above points are number of mature females.....75

Figure 3.9. Proportion of mature female *Platycephalus bassensis* with new POF (< 6 h) at different times of the day during spawning season. Values above bars are numbers of mature females.....76

Figure 4.1. Mean gonad weight ($gr \pm SE$) adjusted for somatic weight of *P. bassensis* at each location and in each season. Means sharing the same letter are not significantly different. Asymmetric SEs resulted from back transformation of logarithmic values. Values in parentheses are sample size.....90

Figure 4.2. Monthly changes in mean gonad weight ($gr \pm SE$) adjusted for somatic weight of *P. bassensis* in three locations. Means sharing the same letter are not significantly different. Asymmetrical SEs resulted from back transformation of logarithmic values. Values in parentheses are sample size.....91

Figure 4.3. Changes in the proportion of mature-sized females in each reproductive stage caught in (a) Tamar River (n= 108), (b) Georges Bay (n= 89), and (c) Coles Bay (n= 232) between March 2001 and May 2003.93

Figure 4.4. Changes in the proportion of mature-sized males in each reproductive stage caught in (a) Tamar River (n= 51), (b) Georges Bay (n= 82), and (c) Coles Bay (n= 142) between March 2001 and May 2003.....94

Figure 4.5. Percentage of sexually mature male and female sand flathead, *Platycephalus bassensis*, by 1-cm length intervals, with a logistic model fitted to the data collected at the height of the spawning season. r^2 for all cases was 0.99. Symbols represent probability of maturity of fish within 1-cm intervals.97

Figure 4.6. Percentage of sexually mature male and female sand flathead, <i>Platycephalus bassensis</i> , by 1-year age classes, with a logistic model fitted to the data collected at the height of the spawning season.....	98
Figure 5.1. The slope (\pm 95% confidence limit) of the total length-somatic weight relationships of mature individuals for each reproductive state. Value above each point is the sample size. r^2 was 0.98 for each state.....	115
Figure 5.2. Residual values for mature <i>Platycephalus bassensis</i> derived from the total length-somatic weight and somatic weight-gonad weight relationships for Coles Bay (triangles) and Georges Bay (squares) at different reproductive states.....	118
Figure 5.3. Average liver lipid concentration of mature (a) females and (b) males <i>Platycephalus bassensis</i> in each reproductive state. Superscripts were generated from a Tukey's HSD post-hoc comparison of means for every combination of sex, season and reproductive state.....	121
Figure 5.4. Average liver water concentration of mature (a) females and (b) males <i>Platycephalus bassensis</i> in each reproductive state. Superscripts were generated from a Tukey's HSD post-hoc comparison of means for every combination of sex, season and reproductive state.....	122
Figure 5.5. The average muscle water content of immature <i>Platycephalus bassensis</i> for every combination of location and season.....	124
Figure 5.6. Liver lipid content vs muscle water content for mature <i>Platycephalus bassensis</i> caught in Coles Bay and Georges Bay.....	125
Figure 6.1. Further clarification of contributing parameters in population structure in <i>Platycephalus bassensis</i> incorporating major finding. [\times] signifies little to no contribution, [\surd] signifies contribution, [?] signifies possible contribution.....	133

TABLE LEGEND

Table 2.1. Collection details of *Platycephalus bassensis* at each location.....17

Table 2.2. Von Bertalanffy growth parameters for *P. bassensis* for each sampling year and each sex from Coles Bay. Values in parentheses are standard errors.....35

Table 2.3. Summary of age-length linear regression, associated with statistics, for *P. bassensis* < 5 years old. Asymmetric CLs resulted from back transformation of logarithmic values.....36

Table 2.4. Estimates of numbers of demographic parameters of *Platycephalus bassensis* from the 3 locations.....39

Table 3.1. The sampling regime and the exact date and time of *Platycephalus bassensis* collection in Georges Bay (GB), Coles Bay (CB), North-West Bay (NWB), and Spring Bay (SB).....53

Table 3.2. Microscopic, macroscopic and histological staging criteria used for sand flathead ovaries.....57

Table 3.3. Macroscopic and histological staging criteria used for sand flathead testis.....58

Table 3.4. Histological staging criteria of atresia in sand flathead ovary.....59

Table 3.5. Percentage of mature female sand flathead in each size category during the spawning season in Coles Bay. Percentage of atresia includes ovaries that are classified in states 2 and 3 of atresia. n = sample size for each size class.....69

Table 3.6. Percentage of mature female sand flathead in each size category during the spawning season in Georges Bay. Percentage of atresia includes ovaries that are classified in states 2 and 3 of atresia. n = sample size for each size class.....70

Table 3.7. Percentage of atresia (sum of states 2 and 3) calculated for each gonad stage for sand flathead caught in Coles Bay. n = sample size for each gonad stage.....72

Table 3.8. Percentage of atresia (sum of states 2 and 3) calculated for each gonad stage for sand flathead caught in Georges Bay. n = sample size for each gonad stage.....72

Table 3.9. Percentage of mature females *Platycephalus bassensis* with and without hydrated oocytes in their ovaries at different times of the day.....76

Table 4.1. Correlations between gonad weight and total length and age for populations in each location.....95

Table 5.1. Months of *Platycephalus bassensis* collection. Values in parenthesis are total number of sampling at each season and location. n = sample size.....111

Table 5.2. ANCOVA exploring the effect of gender, location, and reproductive state on the somatic weight of *Platycephalus bassensis* using total length (TL) as a covariate.....115

Table 5.3. ANCOVA examining the effect of gender, location, and reproductive state on the gonad weight of *Platycephalus bassensis* using somatic weight (SW) as a covariate..... 116

Table 5.4. Geometric mean regression relationships of somatic weight vs gonad weight for mature *Platycephalus bassensis* at different reproductive states with the locations and genders pooled.....116

Table 5.5. Pearson's correlation coefficients between somatic condition and gonad condition in mature <i>Platycephalus bassensis</i> at different reproductive states at each location. See text for details of calculations of somatic and reproductive condition.....	117
Table 5.6. ANCOVA exploring the effect of gender, location, and reproductive state on the liver weight of <i>Platycephalus bassensis</i> using somatic weight (SW) as a covariate.....	119
Table 5.7. ANOVA examining the effect of gender, location, and reproductive state on the muscle water content of mature <i>Platycephalus bassensis</i>	123
Table 5.8. ANOVA examining the effect of location and season on the muscle water content of immature <i>Platycephalus bassensis</i>	123

CHAPTER ONE

GENERAL INTRODUCTION

1.1. POPULATION STRUCTURE

The structure of a fish population is determined by the balance between life-history processes of reproduction, growth, and mortality (Sissenwine, 1984; Beverton & Holt, 1993) (Fig 1.1). Growth and reproduction as two major components of the life history traits are interconnected, energy channelled into the gonads detracts from growth (Roff, 1984). There is thus a direct trade-off between growth and reproduction. The extent of such trade-off in driving population dynamics and the implication of fish condition, at the whole animal level, in quantifying variation in the differential allocation of energy yet to be addressed for fish. This study explore variability in the life history characteristics of sand flathead, *Platycephalus bassensis*, in the context of energy tradeoffs between growth and reproduction. Understanding of energy tradeoffs develops our knowledge in identifying contributing factors responsible for variation of sub-populations.

Given that populations of marine organisms with dispersive larvae recruit into different environments, life history characteristics such as reproduction, growth, and mortality are likely to vary temporally and among sub-populations. Such changes in life history traits will drive the variability in the structure of fish populations (Ihssen *et al.*, 1981). Changes in population structure, in turn, may alter life history traits (Elliott, 1987; Clutton-Brock *et al.*, 1997; Rodd & Reznick, 1997). Despite the general recognition that life history traits are the fundamental component of population dynamics (Cole, 1954; Sinclair, 1995), few studies have assessed the relationship between life history traits and population response (Begg *et al.*, 1999; Leips & Travis, 1999). Of those that do study the relationship between life history traits and population dynamics, almost none assess the variability among populations

over the time and the possible consequence of such variation in population structure. The rarity of studies that examine variability in population structure of a species limits our ability to identify general mechanism(s) underlying changes in population structure.

Apart from influences of reproduction, growth, and mortality, behavioural process such as migration or movement of fish between areas may drive changes in population structure (Jennings & Kaiser, 1998). Such variations in population structure are often unpredictable and cause uncertainty for the fisheries scientist.

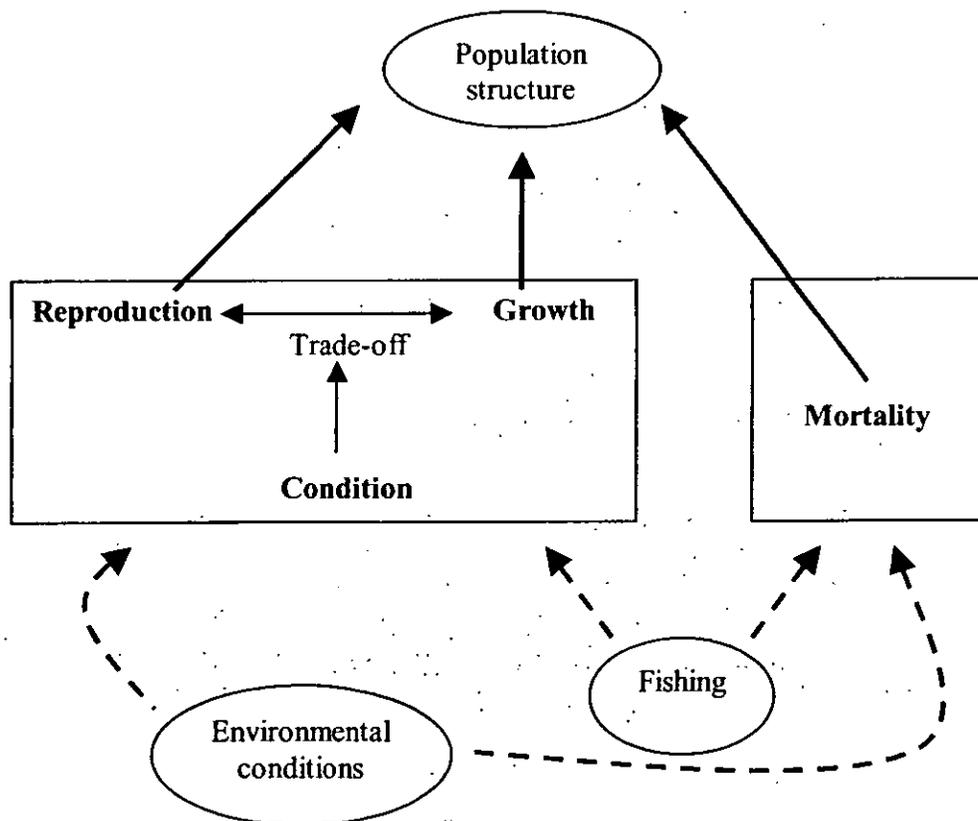


Figure 1.1. Schematic of population formation and contributing parameters in this process.

Population growth is a result of recruitment of new animals to the population and the growth of individuals already in the population (Beverton & Holt, 1993; Haddon, 2001). Since individuals of similar size can have vastly different ages, the size structure of a population alone is generally unable to reveal underlying processes of population growth. However, size-at-age data that can be used to describe patterns of growth and to estimate growth rates of individuals revealing the process of increase in stock biomass (Kirkpatrick, 1993; Haddon, 2001). Additionally, estimates of growth rate, together with size/age structure and longevity, are needed to estimate rates of population turnover and their resilience to different mortality regimes (Choat *et al.*, 2003). Apart from growth, reproduction is central to the capacity of a population to increase in size. In unfished populations, variability in reproductive potential may indicate local adaptation, either through genetically isolated demes or through responses to a broad range of physical and biological conditions (Jennings & Beverton, 1991; see review in Stearns, 1992).

Although the implications of growth and reproduction in population structure and its variability among populations is unquestionable, these two parameters are tightly interconnected (Wootton, 1979; Roff, 1984). Energy channelled to reproduction potentially affects growth and survival. For fish in general, a 'growth cost' of reproduction is evident in the asymptotic growth curve where growth slows after maturation (Roff, 1984; Kozlowski & Teriokhin, 1999). If energy is limiting, such trade-off is certainly reasonable (Roff, 1992; Stearns, 1992). Alternatively, it is possible that energy is not limiting and growth rate is not limited by input (Roff, 1992), thereby trade-off may not occur. Theories on resource allocation between growth and reproduction remain almost untested in practice for most animals (Heino & Kaitala, 1999), because *separating the effect of exogenous factors on life history*

traits is difficult. For fishes our understanding of trade-off between reproduction and growth is far from complete and limited to a few studies (Roff, 1982; Reznick, 1983). As a result, spatial variability in growth and reproduction in the context of energy allocation has not been explored. Theoretically, spatial differences in the pattern of energy allocation may be seen in spatial variation in growth rates, especially where differences in the level of reproductive investment are also evident. Such variation in growth rate and reproductive output can drive population dynamics. Therefore, examination of spatial variability in somatic condition highlights the implication of energy allocation in driving growth and reproduction processes.

The body size of a fish species determines much of its ecology, life history, and vulnerability to the exploitation (Charnov, 1993; Pope *et al.*, 1994; Kerr & Dickie, 2001). Life history traits such as fecundity, size/age at maturity, and maximum size/age are strongly correlated to body size (Reynolds *et al.*, 2001; Hutchings, 2002). There is a significant correlation between the life history traits of size/age at first reproduction, mortality, and growth rates in local populations (Roff, 1984). Life history theory predicts that populations with high adult mortality rates will select for individuals that mature earlier in their lifetime, thereby increasing reproductive effort in younger age classes (Schaffer, 1979; Stearns, 1983; Charlesworth, 1994). However, as fecundity is proportional to the size/age of fish (Bagenal, 1966; Kjesbu *et al.*, 1998) small individuals would not be able to invest significantly in egg production. The trade-off between current and future reproduction (Roff, 1992; Stearns, 1992; Rochet *et al.*, 2000) suggests that the lower fecundity at lower size/age maturity, would be compensated through a higher fecundity at larger size/age. Maturing at a greater age/size results in lower mortality (Roff, 1981;

Rochet *et al.*, 2000) and because of higher fecundity of larger individuals, recruitment of new animals to the population is expected to be higher. Variation in life history traits has the potential to drive population dynamics, therefore investigations of such variability is central to understanding the mechanisms underlying such dynamics.

Life history traits such as growth, fecundity, and size/age at maturity may vary among populations as a result of variations in the environment that an animal lives (Vollestad & Labee-Lund, 1990; Jennings & Beverton, 1991). For example, growth rates of fish will typically increase with temperature (Atkinson, 1994), with faster growth rates decreasing the age at maturity (Trippel, 1995; Poortenaar *et al.*, 2001). Fish growth is also highly sensitive to resource availability, and at low densities where intraspecific competition is expected to be reduced fish growth rate can be high (Borisov, 1978; Frank & Leggett, 1994; Rijnsdorp, 1994). Because of the positive relationship between fecundity and fish size (Bagenal, 1966; Kjesbu *et al.*, 1998), any environmental factor that influence growth rate and body size may indirectly effect fecundity and reproductive potential (Reiss, 1989).

The size and age structure of a population can also be the result of size selective fishing mortality (Rothschild, 1986). Spatial variability in mortality regimes of different size/age classes can cause variations in the size and age structure of populations (Law & Rowell, 1993; Bertschy & Fox, 1999). Greater rates of mortality of larger and older individuals, which are generally part of the spawning stock, directly affects the reproductive potential of a population (Garrod & Horwood, 1984; Rijnsdorp *et al.*, 1991; Begg & Marteinsdottir, 2003). Determination of variability in fish population dynamics is important and necessary to fisheries management for the development of optimal harvest and monitoring strategies

(Smith *et al.*, 1990; Law, 2000). Many studies have documented differences in population parameters among species (Turchin & Taylor, 1992; Clutton-Brock *et al.*, 1997), but relatively few have examined spatial differences in exploited populations of a single species (Trippel, 1995), or populations in distinct habitat types (Leips & Travis, 1999). This study examines population dynamics of a fish species at different locations where recreational fishing pressure occurs in all locations.

Although over the last few decades, recreational fisheries has grown in coastal waters all over the world (Radomski *et al.*, 2001; Post *et al.*, 2002), it is still regulated without specific management plans (Pereira & Hansen, 2003). Even if a recreational fisheries management plan exists it does not usually consider temporal and spatial variability in population structure. Almost all fisheries management is based on assumption that populations of a fish species are uniform at least in the regional scale. Such management, together with a common view on self-sustaining of recreationally fished stocks or underestimated impact of recreational fishing, may fail to prevent long-term effects of recreational fishing and can end to an invisible collapse (Post *et al.*, 2002; Radomski, 2003). Responses of different populations to exogenous factors (e.g. fishing pressure, patterns of fish movement, and environmental conditions) and their resilience to such factors is now a concern of fisheries managers, but poorly documented. Quantifying the nature and magnitude of differences in life history parameters of sand flathead, *Platycephalus bassensis*, as an example of a fish species that is under recreational fishing pressure (Kailola *et al.*, 1993; Lyle & Campbell, 1999), allows potential mechanisms of plasticity in population to be identified. This study will quantify variability in demographic parameters and reproductive potential among sub-populations, and therefore determine the condition of each population. Such information is required to progress

spatial and temporal management of the recreational fishery for this species and will assist the application of these approaches to other species.

1.2. GENERAL OBJECTIVES

The general aim of this study was to examine spatial and temporal variation in life history characteristics of the sand flathead, *Platycephalus bassensis*. This was achieved through intensive field sampling and an *in vitro* experiment with particular emphasis on identifying the spatial and temporal variability in growth, reproduction, and condition of adult population.

1.3. CHAPTER SCOPES

This thesis consists of four data chapters; each one comprising a stand-alone manuscript for publication, therefore there may be areas in the text that are slightly repetitive.

Chapter 2: Spatial and temporal variation in population dynamics of sand flathead, *Platycephalus bassensis*.

Through two years of sampling this chapter's primary aim was to quantify spatial differences in the population structure of sand flathead. Size/age composition, patterns and rates of growth and mortality rates were quantified for each population. The growth rate in the first year of life was also assessed using the mean radius of the otolith first increment. A snapshot of monthly changes in water temperature and salinity was used as an index for environmental conditions in each location.

Chapter 3: Reproductive biology of sand flathead, *Platycephalus bassensis*.

Histological analysis of ovary and testis was used to quantify reproductive characteristics of sand flathead. With the combination of acquired information from field sampling and results from an *in vitro* experiment this study aimed to investigate the dynamics of spawning of sand flathead and differences in reproductive strategy. Lunar and diel spawning periodicity and spawning frequency were determined based on the presence of postovulatory follicles in the ovary. Additionally, the presence and extent of atresia in the ovary was used to quantify the proportion of spawning in different length classes during the spawning period.

Chapter 4: Spatial and temporal variability in reproductive ecology of sand flathead, *Platycephalus bassensis*.

Based on the reproductive biology of sand flathead, this chapter describes spatial and temporal differences in reproductive life history characteristics. This study highlighted the variability in gonad weight and its correlation to body size that was considered a determinant of the reproductive output of each population. Size and age at maturity, as one of the most essential components for understanding of life history strategy, was also estimated for each population.

Chapter 5: Spatial variation in condition and patterns of repro-somatic investment in sand flathead, *Platycephalus bassensis*.

To understand the process and patterns of resource allocation between reproduction and other competing needs such as maintenance and growth, somatic, reproductive, and liver conditions were assessed for mature and immature individuals separately. For mature fish, condition was examined in the context of

reproductive state; pre-spawning, spawning, and post-spawning. Proximal analysis was also carried out for further investigation on process of energy partitioning. Lipid and water contents of liver and muscle water content was measured for mature and immature individuals separately.